# Supplementations of *Bacillus* spp. on gut microbiota of weaned pigs under diarrheal stress

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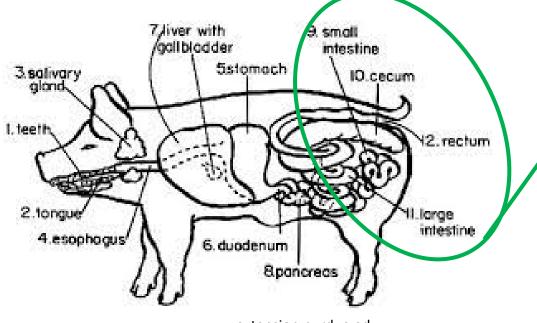
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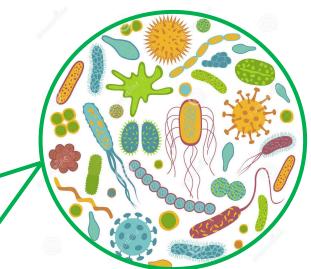
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## Gut microbiota

 The digestive system (gut) is crucial for nutrient metabolism and immunity regulation in pigs



extension.purdue.edu



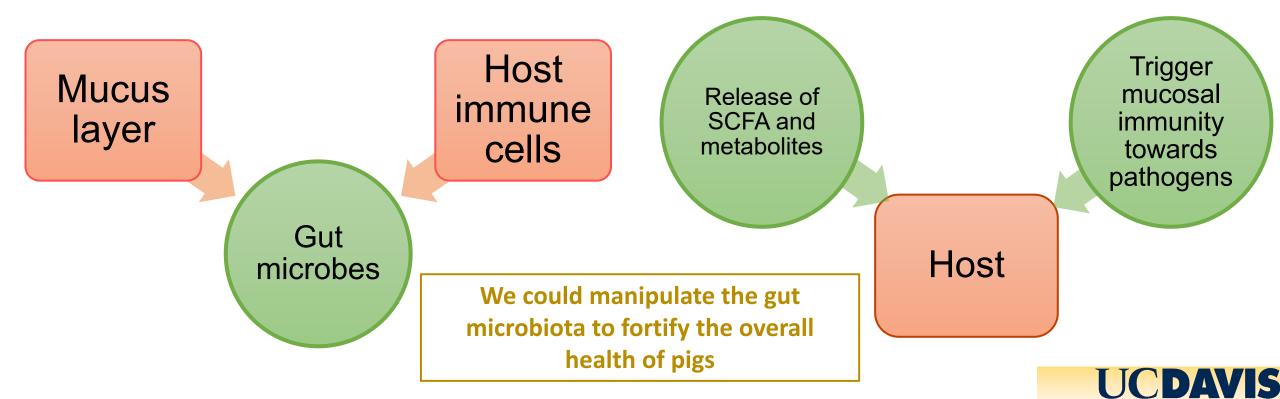
 Microbes are to harbor innumerably in the gut and have been overlooked to understand their benefits for the pig (host)

 The dynamic and diverse population of microbes in the gut is collectively referred to as the <u>gut</u> <u>microbiota</u>



#### Gut microbiota

 Microbes and their host holds a symbiotic relationship, in which the host allows microbes to habituate in the intestines while the microbes benefits the host's health



#### Post-weaning stress

- Weaning becomes one of the most stressful moments in pigs
  - Separation from sow and possibly their littermates
  - Change of environment
  - Diet change
- Weaning stress reduces feed intake, which can cause a cascading effect toward pig's health
  - Lower feed intake = less metabolizable energy = disturbance of intestinal development in growing pigs

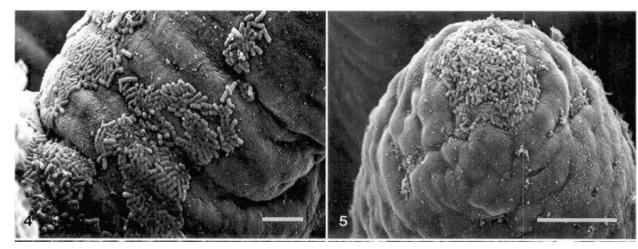


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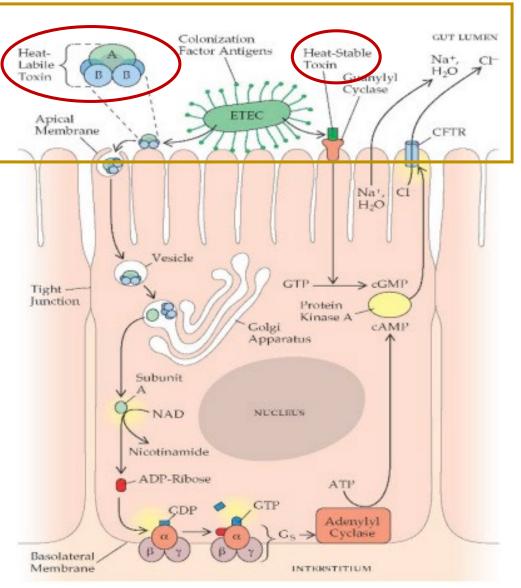
#### **ETEC** pathogenesis

## Post-weaning diarrhea

- Post-weaning stress can cause diarrhea
- Post-weaning diarrhea is commonly induced by enterotoxigenic *E. coli* (**ETEC**)
- Leads to high morbidity rate in weaning pigs along with huge economical loss in the swine industry



Vet Pathol 29:239-246 (1992)



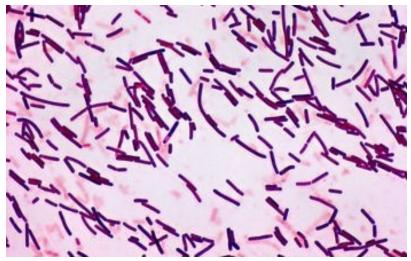
Microbial pathogenesis 117: 162-169 (2018)



### Bacillus spp.: Potential feed additives

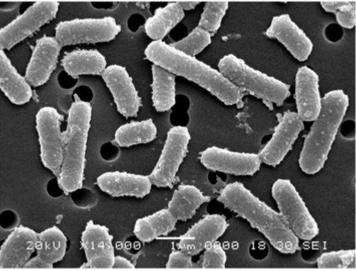
• Our previous study (He et al., 2020) observed that supplementing *B.* subtilis reduced diarrhea and enhanced growth performance of weaned pigs experimentally infected with ETEC

microbe-canvas.com



Bacillus subtilis

Annals of clinical microbiology and antimicrobials, 14(1), 1-11.

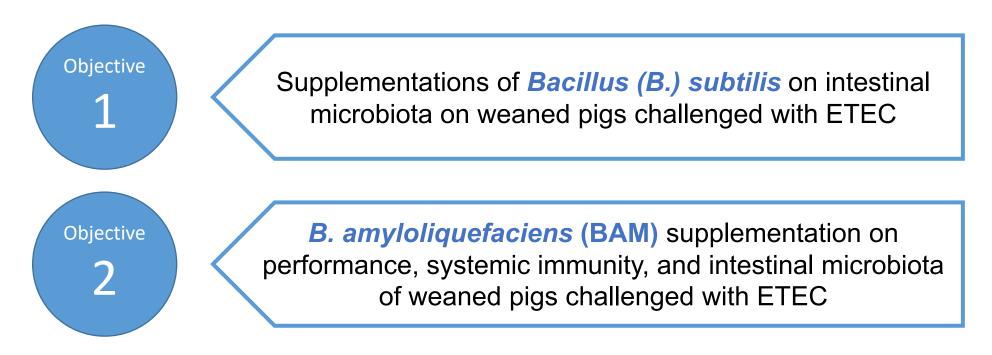


Bacillus amyloliquefaciens

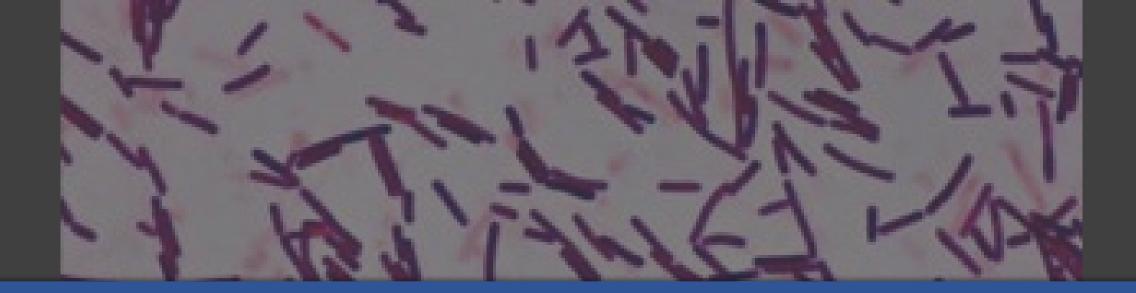


## Bacillus spp.: Potential feed additives

• However, impact of *Bacillus* spp. on gut microbiota of weaned pigs under post-weaning diarrhea is not yet understood

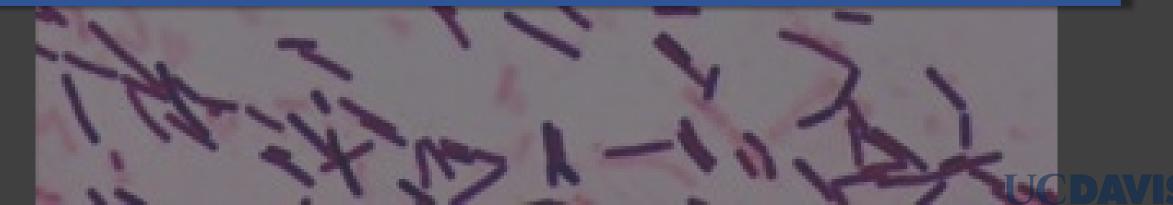






# Supplementations of *B. subtilis* on intestinal microbiota on weaned pigs challenged with ETEC

Objective 1



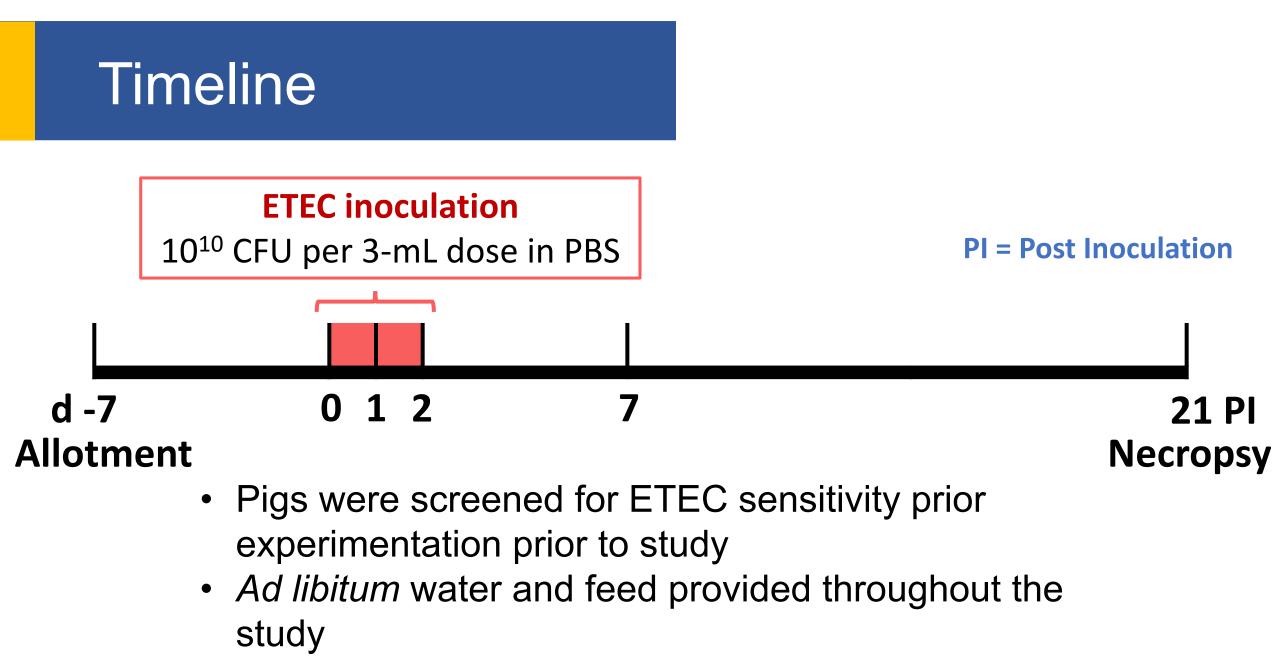
### Materials & methods

- 48 weaned pigs (6.17 ± 0.36 kg)
  - Around 21-24 day of age
- 4 treatments (12 pigs / treatment)

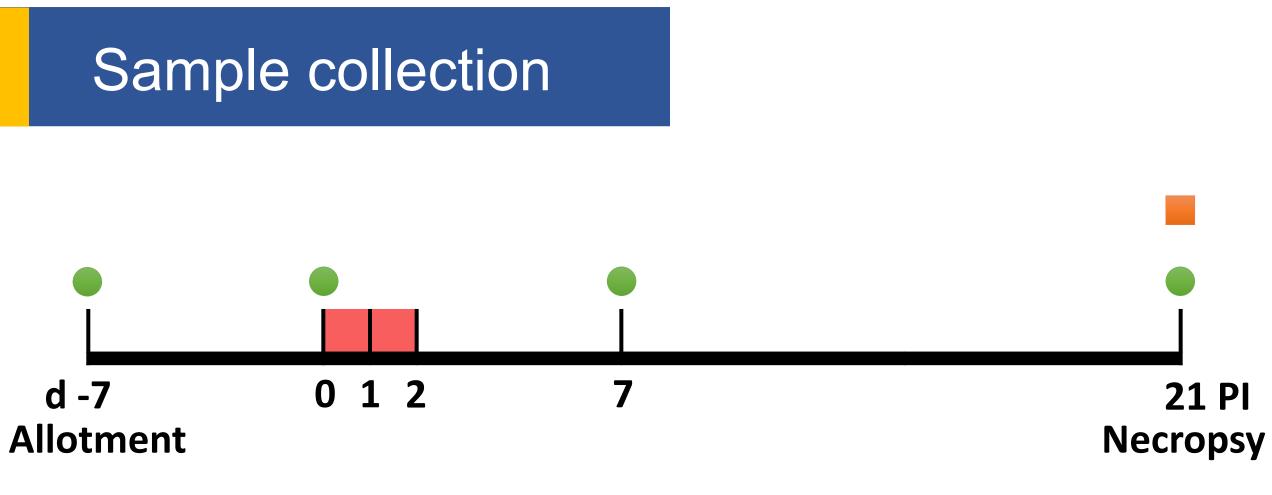
Sham	NC	Negative control with basal diet		
ETEC	PC	Positive control with basal diet		
	AGP	Basal diet with 50 mg/kg carbadox		
	DFM	Basal diet with 500 mg/kg B. subtilis		

AGP = antibiotics growth promoter (carbadox) DFM = Direct fed microbials (*B. subtilis*)





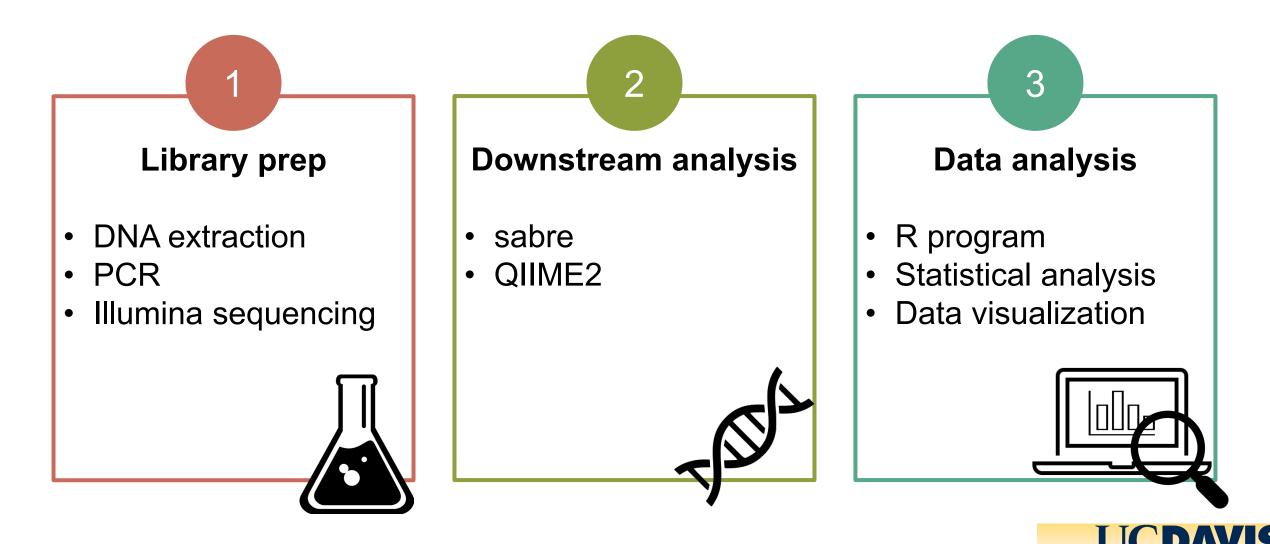




# Fecal samples collected Jejunal digesta, ileal digesta, and cecal content



# 16S rRNA sequencing



#### Gut microbiota analysis

#### Alpha diversity:

- Shannon index (richness & evenness)
- Chao1 index (richness only)

**Beta diversity:** 

Bray-Curtis dissimilarity

#### **Taxonomic analysis:**

 Relative abundance in phylum and genus levels

# Statistical analysis

#### Alpha diversity and relative abundance

- <u>Normal distribution</u> = ANOVA followed by estimated marginal means (EMMEANS)
- <u>Non-normal distribution</u> = Kruskal-Wallis followed by Conover test
- Beta diversity
  - Betadisper followed by adonis function using vegan package in R

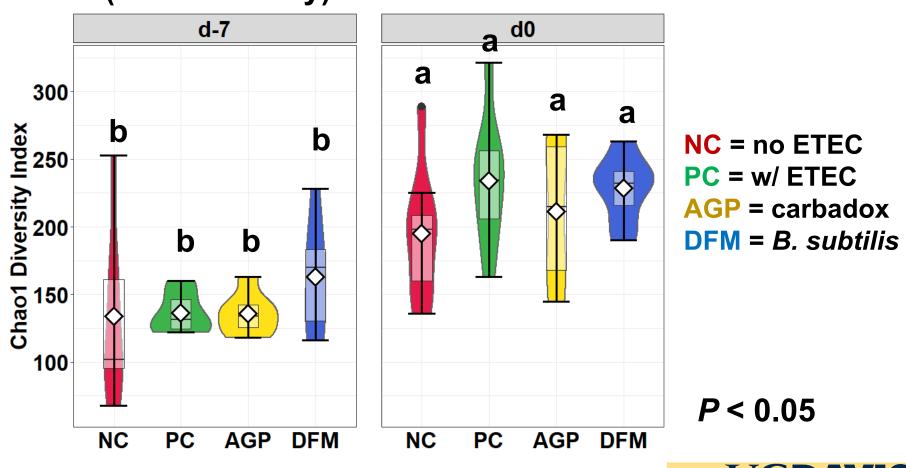


# Results: Fecal microbiota

No significant difference observed in Shannon diversity (richness & evenness)

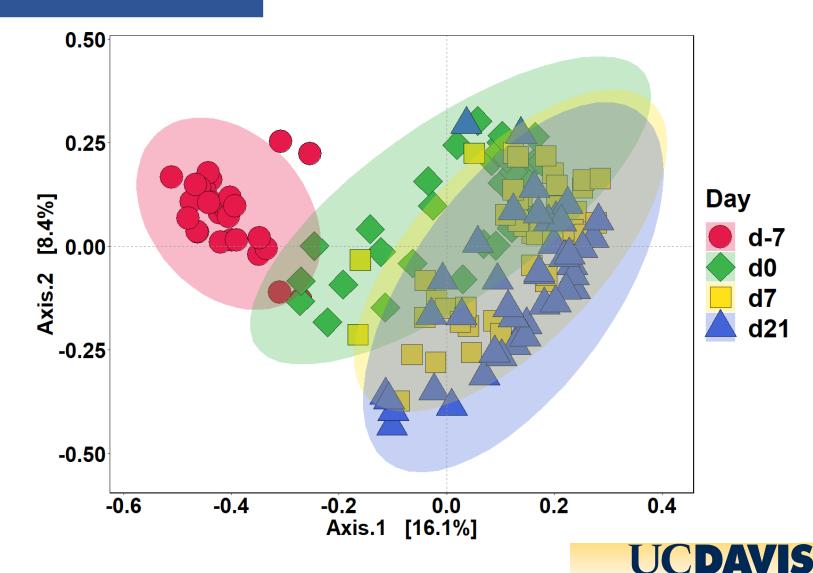
Alpha diversity: Chao1 index (richness only)

Chao 1 index was
 increased between
 d -7 and d 0 for all
 dietary treatments

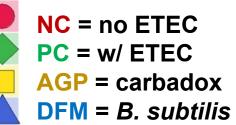


### Feces: Beta diversity

- Each point represent a sample
- Color and shape represent day sampled
- The farther the point is from another point, the more likely there is difference in microbial composition between the 2 points

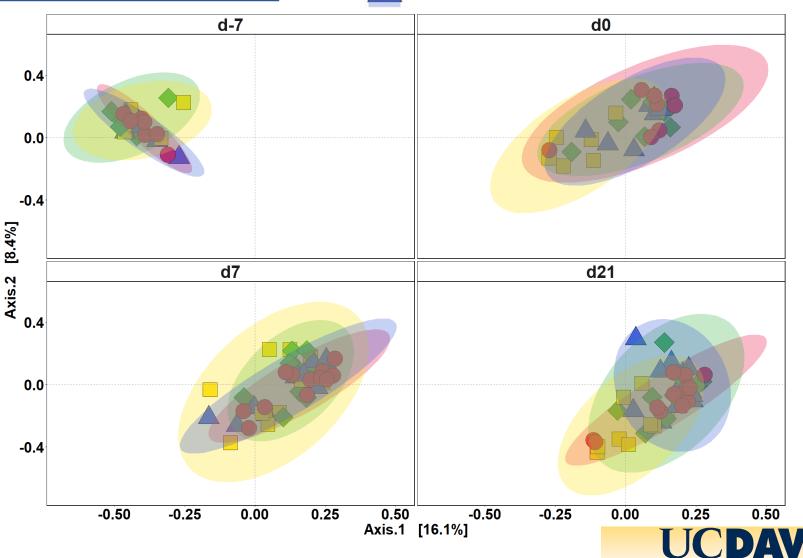


### Feces: Beta diversity



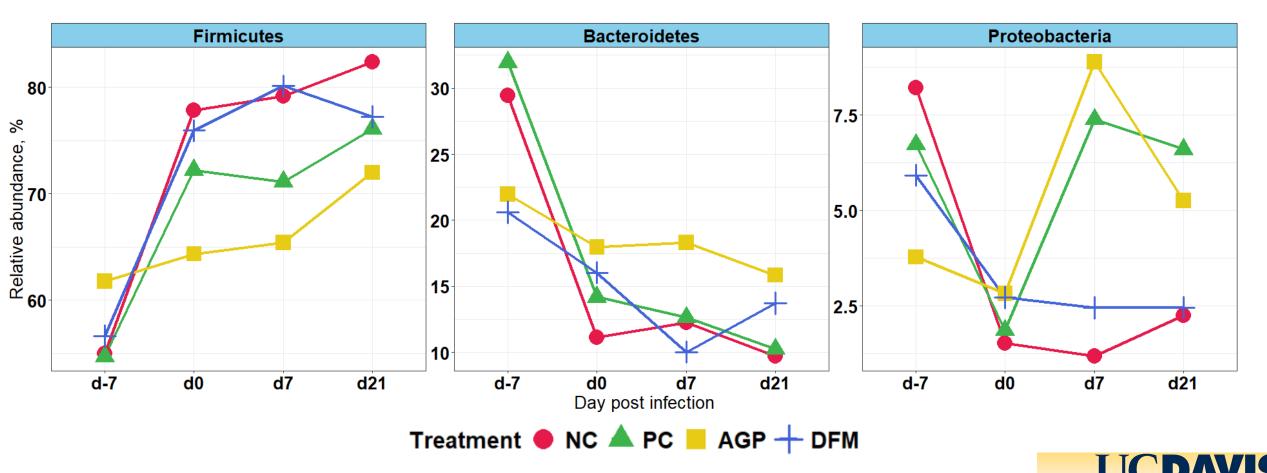
#### **Treatment\*Day interaction**

- Facetted by sampling days
- Color and shape
  represent treatment



# Feces: Phylum

- Firmicutes increased over time
- Bacteroidetes decreased over time
- Proteobacteria remained the same

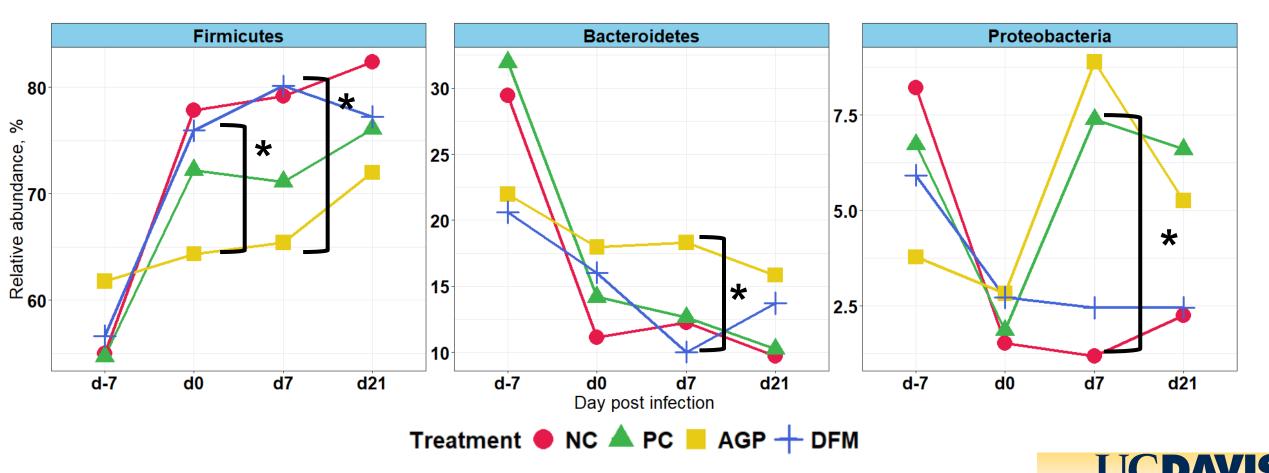


#### **Relative abundance: Phylum level**

# Feces: Phylum

**Relative abundance: Phylum level** 

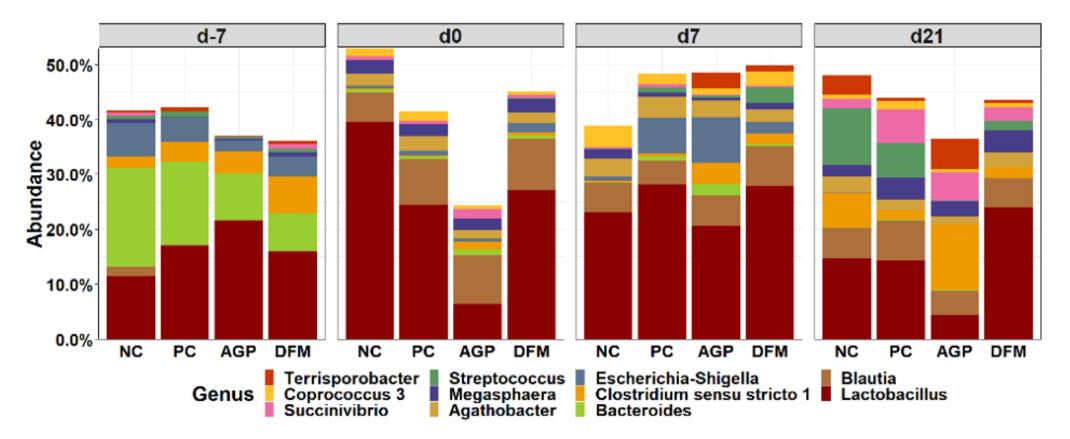
- Firmicutes increased over time
- Bacteroidetes decreased over time
- Proteobacteria remained the same



#### Feces: Genus

 Lactobacillus was greater in DFM than in AGP on d 0 and 21 PI

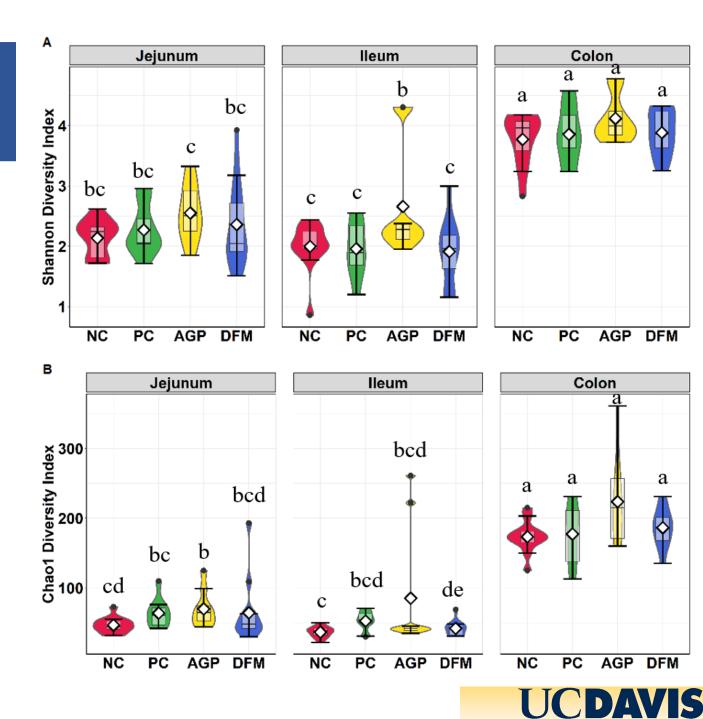
#### **Relative abundance: Genus level**



### Results: Intestines

#### Alpha diversity: Intestinal sites

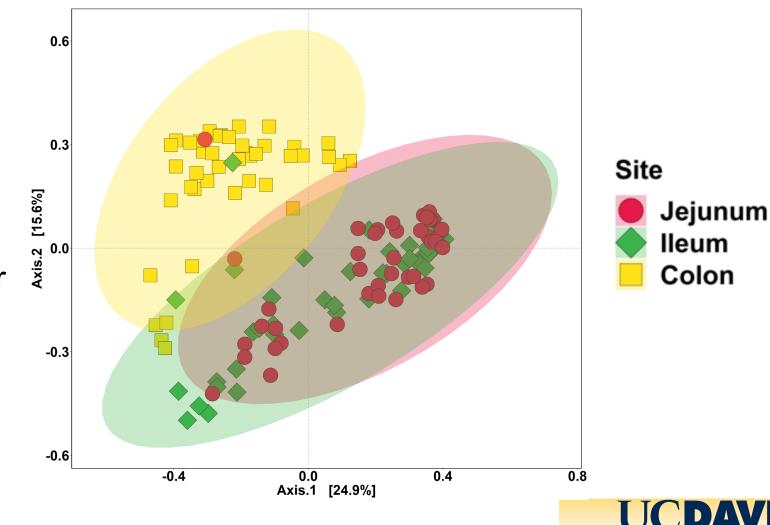
- Samples were collected on d 21 PI
- Shannon and Chao1 indices were greatest in colon



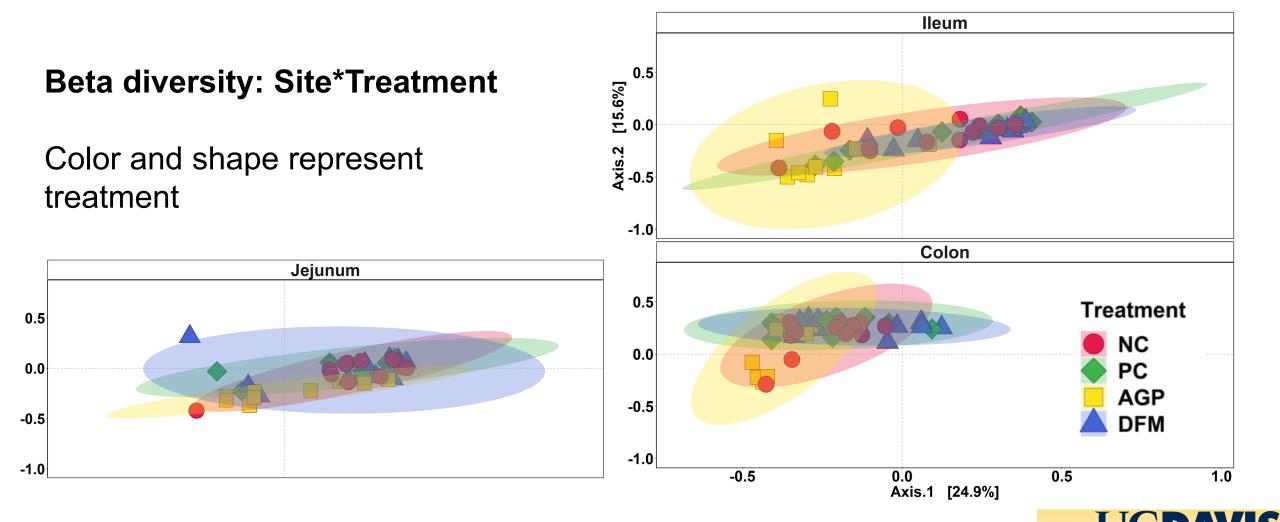
### Intestines: Beta diversity

#### **Beta diversity: Intestinal sites**

Color and shape represent intestinal site (jejunum, ileum, or colon)



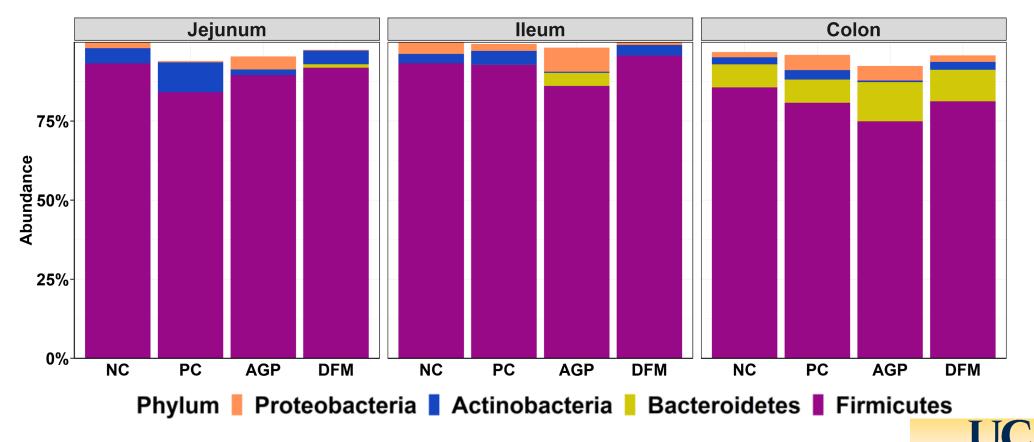
### Intestines: Beta diversity



# Intestines: Phylum

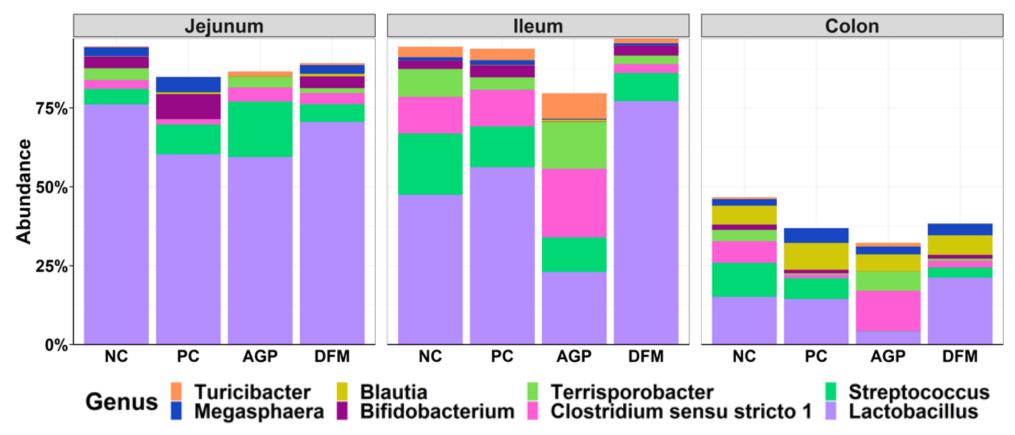
#### **Relative abundance: Phylum level**

- Actinobacteria was greater in ileal digesta from PC than from AGP
- Bacteroidetes was greater in AGP than in DFM in ileal digesta
- Proteobacteria was greater in AGP than in DFM in jejunal and ileal digesta



### Intestines: Genus

 Lactobacillus and Bifidobacterium were greater in DFM than AGP and PC in jejunum, ileum, and colon on d 21 PI

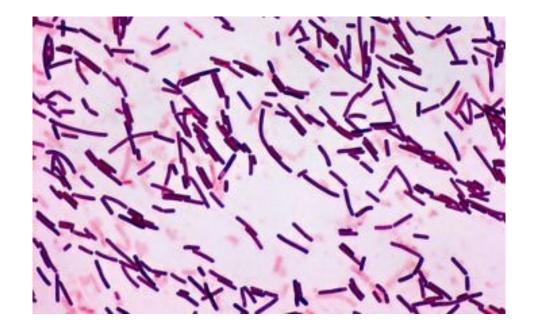


#### **Relative abundance: Genus level**

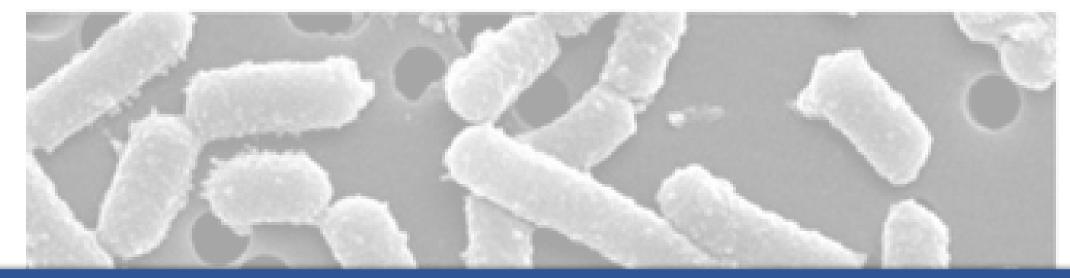


## Conclusion

- Age, intestinal sites, ETEC, and diet all contributed to the modulation of gut microbiota
- Carbadox supplementation increased relative abundance of gram-negative bacteria including Bacteroidetes and Proteobacteria
  - Could increase risk of antibiotics resistance
- B. subtilis supplementation was associated with increase of beneficial microbes in the intestinal microbiota of weaned pigs under ETEC challenge







*B. amyloliquefaciens* (BAM) supplementation on performance, systemic immunity, and intestinal microbiota of weaned pigs challenged with ETEC

Objective 2

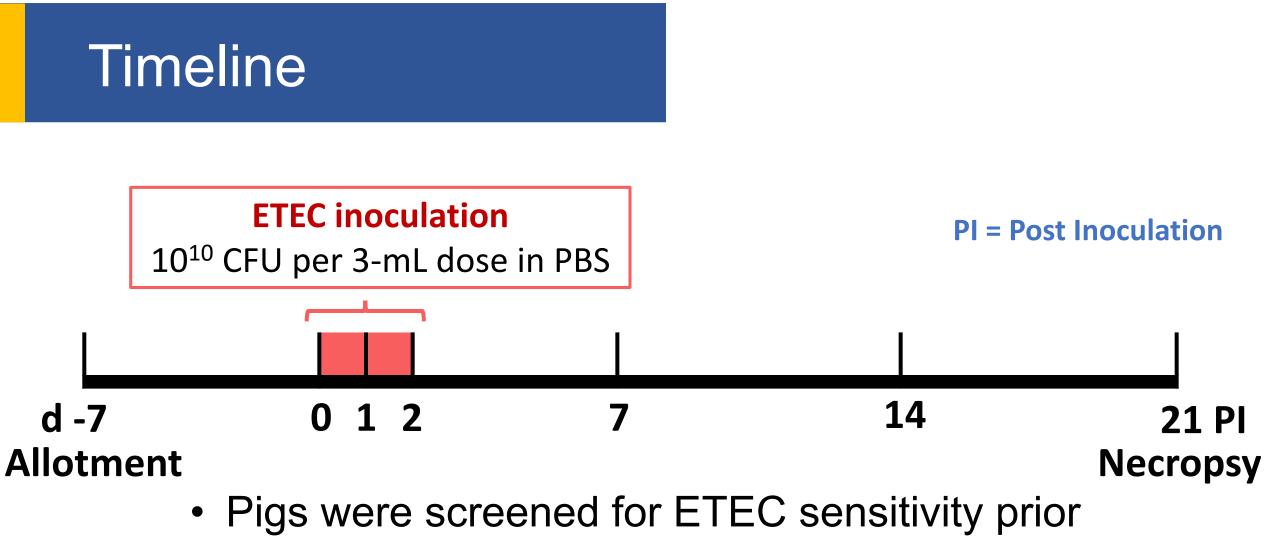


### Materials & methods

- 50 weaned pigs (7.41 ± 1.35 kg)
  - Around 21-24 day of age
- 5 treatments (10 pigs / treatment)

Sham (-)	CON -	Control diet		
	BAM -	0.10% inclusion rate with 10 <sup>9</sup> CFU/kg BAM		
ETEC (+)	CON +	Control diet		
	BAM +	0.10% inclusion rate with 10 <sup>9</sup> CFU/kg BAM		
	AGP +	50 mg/kg of Carbadox		





- to experimentation
- Ad libitum water and feed provided

#### Measurements

#### **1. Growth performance**

- Bodyweight
- Average daily gain (ADG)
- Average daily feed intake (ADFI)

#### 3. Systemic immunity

Total and differential blood cell count

#### 2. Diarrhea frequency

 Diarrhea score (1 = normal feces, 5 = watery diarrhea)

#### 4. Fecal and ileal microbiota

- Alpha and beta diversity
- Relative abundance



# Statistical analysis

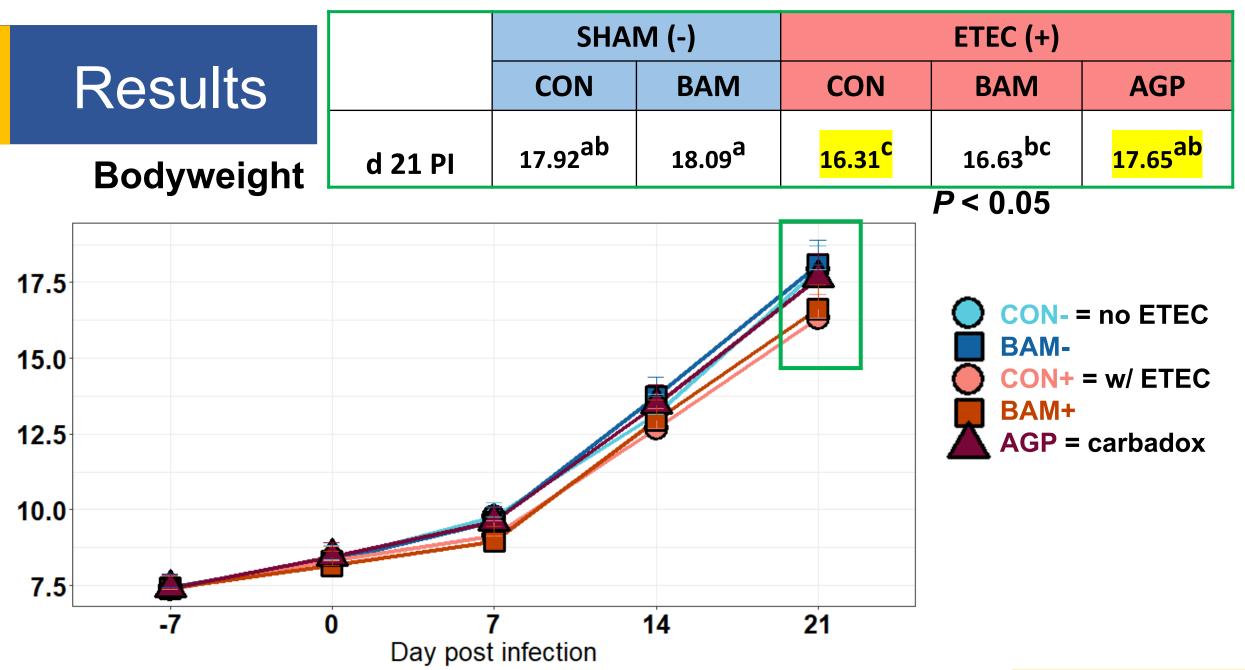
#### PROC MIXED of SAS

- Randomized complete block design
- <u>Pig</u> as experimental unit
- Diet and challenge as main effect
- Block as random effect

# Statistical analysis: Gut microbiota

- Alpha diversity and relative abundance
  - <u>Normal distribution</u> = ANOVA followed by estimated marginal means (EMMEANS)
  - <u>Non-normal distribution</u> = Kruskal-Wallis followed by Conover test
- Beta diversity
  - Betadisper followed by adonis function using vegan package in R

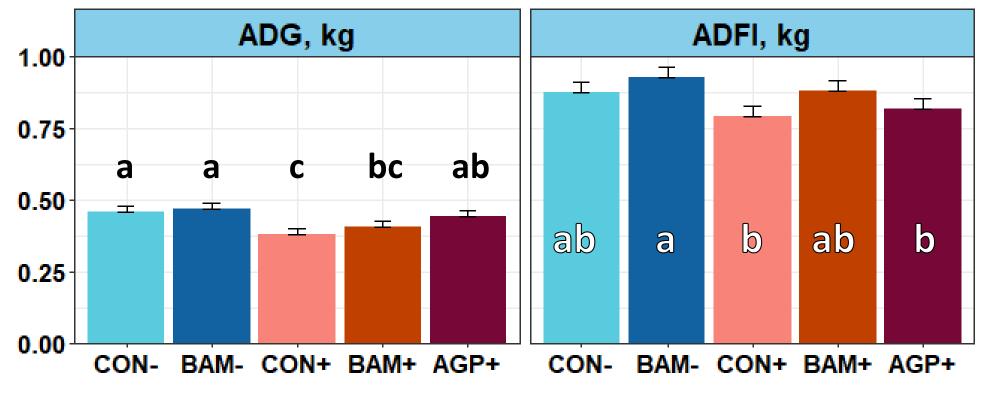




UCDAVIS

#### Growth performance

#### d0 to d21 Pl



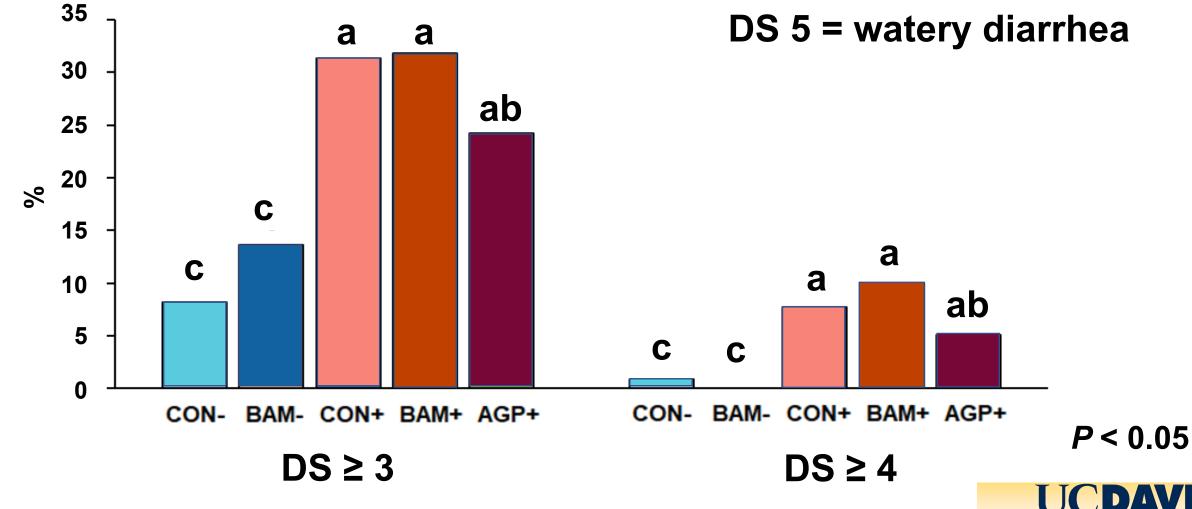
*P* < 0.05



# Frequency of diarrhea

DS = Diarrhea score

#### DS 1 = normal feces **DS 5 = watery diarrhea**



### WBC counts

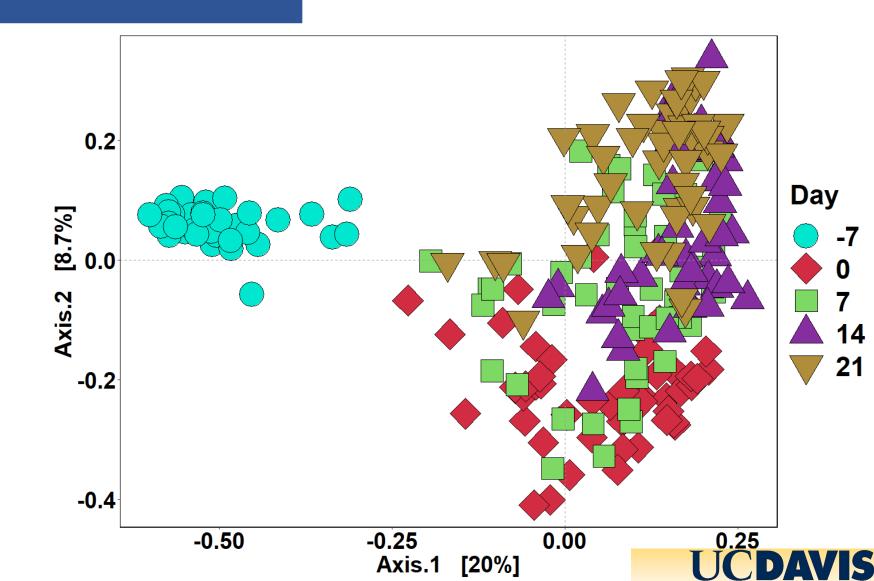
*P* < 0.05

Day Pl	SHAM (-)		ETEC (+)			
	CON	BAM	CON	BAM	AGP	
Lymphocyte, 10 <sup>3</sup> /µL						
0	4.86 <b>b</b>	4.75 <sup>b</sup>	5.23 <sup>b</sup>	6.31 <b>ab</b>	7.14 <sup>a</sup>	
7	5.28 <b>b</b>	6.21 <b>ab</b>	8.71 <sup>a</sup>	8.26 <sup>a</sup>	8.22 <sup>a</sup>	
21	5.59 <b>b</b>	5.63 <b>b</b>	7.30 <sup>a</sup>	5.36 <b>b</b>	6.79 <b>ab</b>	
Neutrophil, 10 <sup>3</sup> /µL						
14	7.62 <sup>b</sup>	7.22 <b>b</b>	9.49 <sup>a</sup>	7.92 <b>b</b>	7.27 <b>b</b>	
	ah	ah		bc		

#### Fecal microbiota

**Beta diversity** 

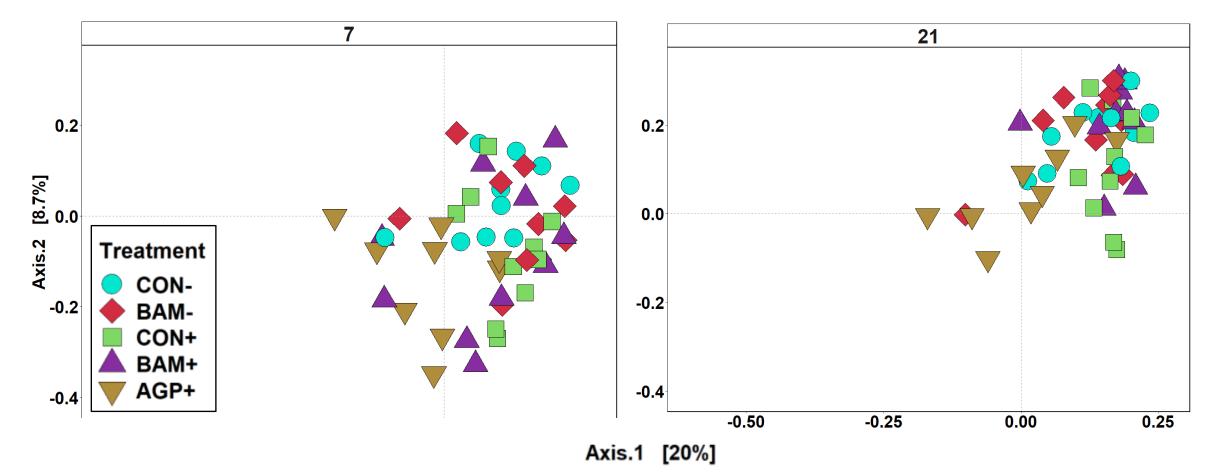
Color and shape represent sampling day



#### Feces: Beta diversity

#### Treatment\*Day

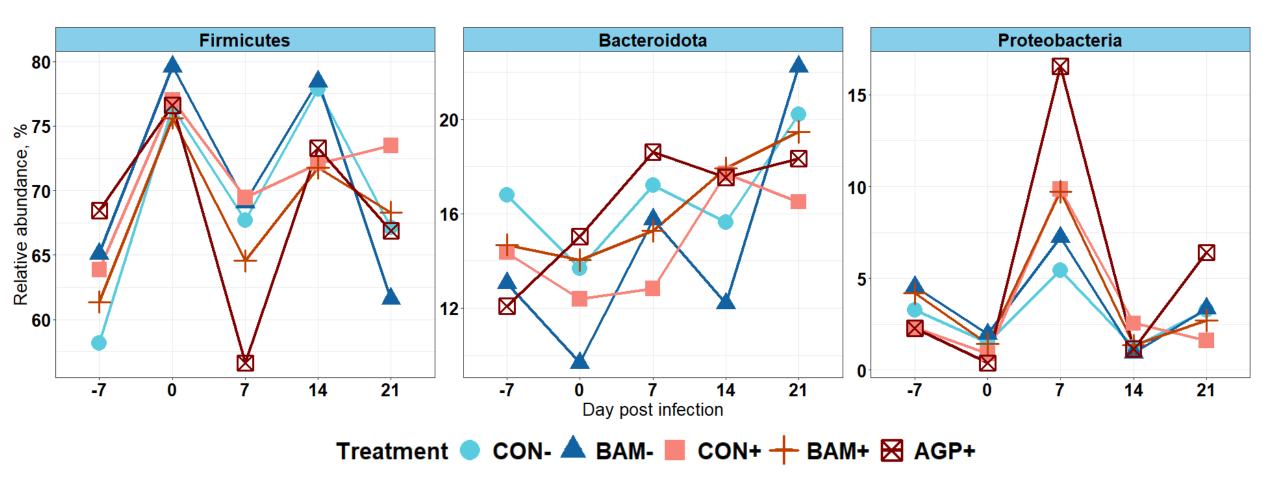
• AGP+ samples were clustered together





#### Feces: Phylum

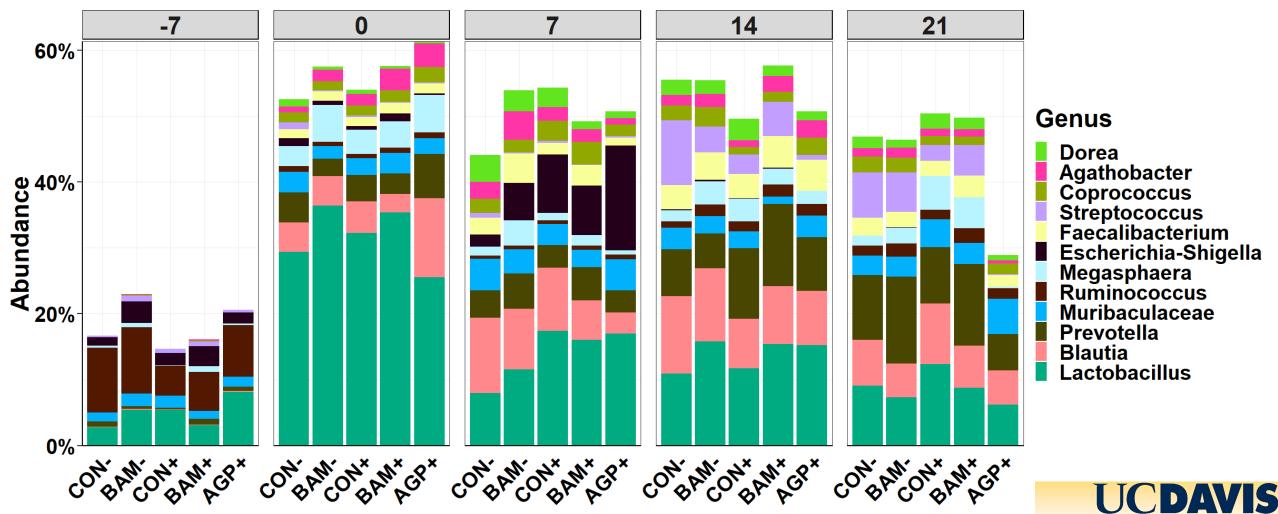
Firmicutes decreased between d 0 and 7 PI in AGP+ Proteobacteria increased between d 0 and 7 PI in CON and AGP





#### Feces: Genus

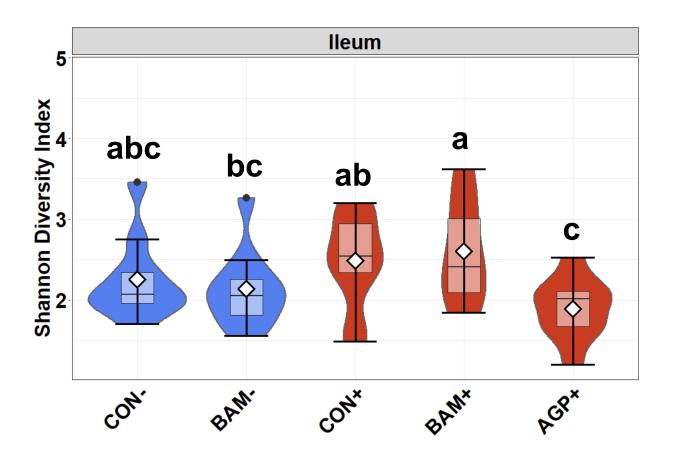
- Lactobacillus was greater in CON+ than CON- on d 7 PI
- Blautia and Prevotella was greater in AGP+ than in BAM+ on d 0
- Prevotella was greater in BAM+ than AGP+ in d 21 PI



#### lleal microbiota

#### Alpha diversity (Shannon)

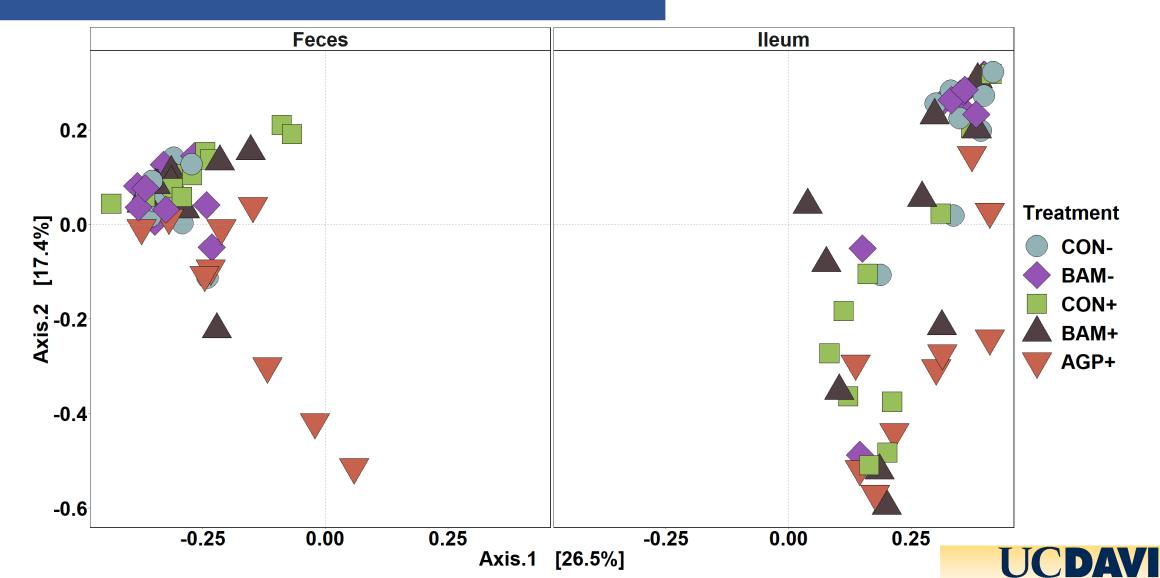
- Ileal digesta collected on d 21 PI
- CON+ and BAM+ had greater Shannon diversity index than AGP+ in ileum





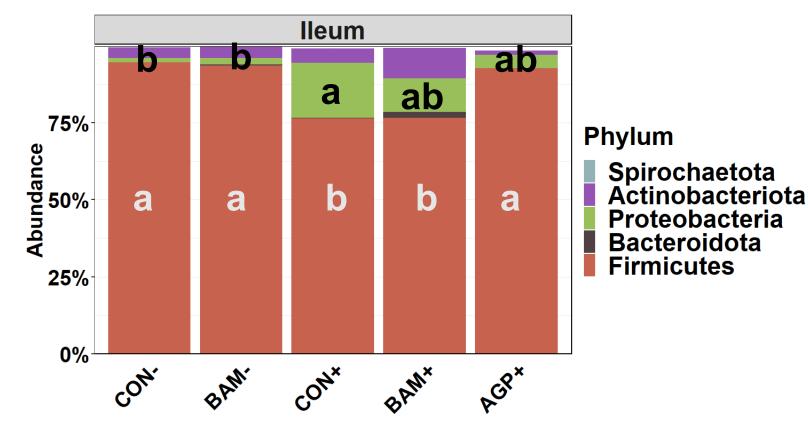
#### **Ileum: Beta diversity**

Separated clusters in ETEC infected and non-infected treatment groups in ileal digesta



# lleum: Phylum

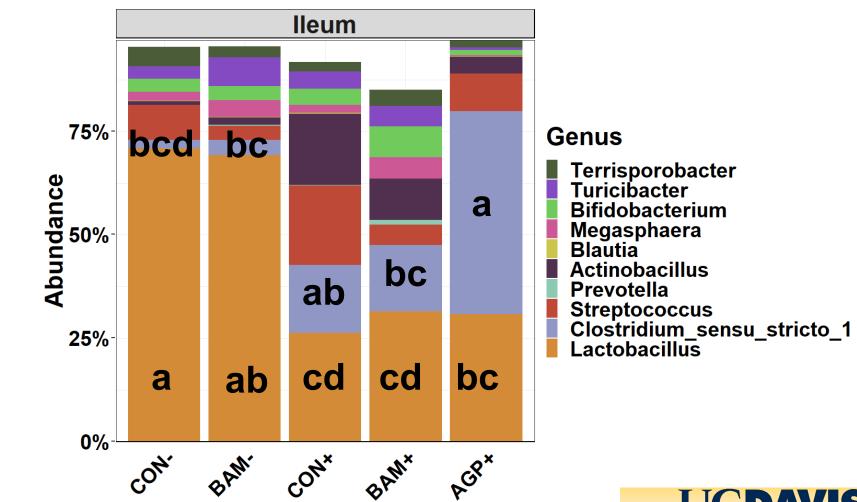
- Firmicutes was greater in AGP+ than in CON+ and BAM+ in ileal digesta
- Proteobacteria was greater in CON+ than in CON-





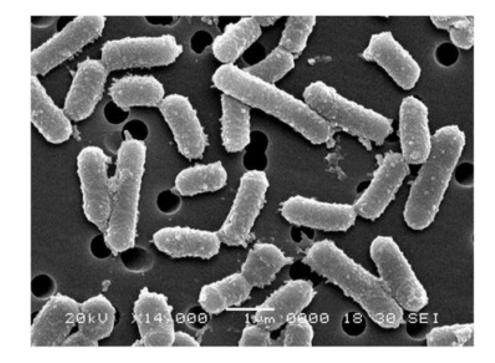
#### lleum: Genus

- Lactobacillus was greater in CON- than in ETEC infected groups
- Clostridium sensu stricto
  1 was more abundant in
  AGP+ than BAM+ in ileal
  digesta



# Conclusions

- *B. amyloliquefaciens* supplementation to weaned pigs challenged with ETEC
  - Tended to enhance growth performance
  - · Had limited effects on diarrhea
  - Reduced systemic inflammation
  - Altered fecal and ileal microbiota differently from carbadox supplementation
- B. amyloliquefaciens solely may not provide weaned pigs with growth enhancement and acute diarrheal alleviation as similarly as carbadox



# **Bacillus** spp. can vary in how they are used how they impact different animal species

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