Nutritional intervention for the intestinal development and health of weaned pigs

Yanhong Liu

Department of Animal Science University of California, Davis, CA

DSM Science & Technology Award Symposium UCDAVIS

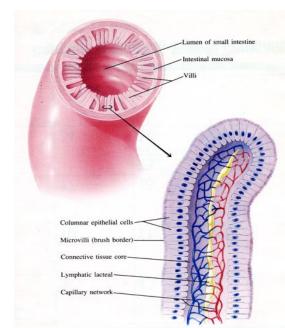
Outline

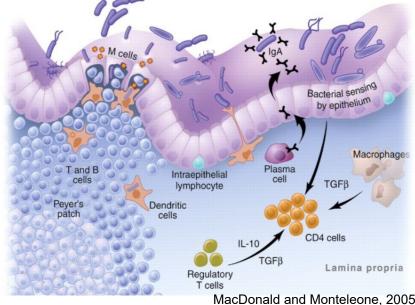
- Weaning stress on intestinal development and health
- How to define a healthy gut
- Nutritional intervention
 - Functional amino acids
 - Short chain fatty acids
- Take home message



Focus on the GUT

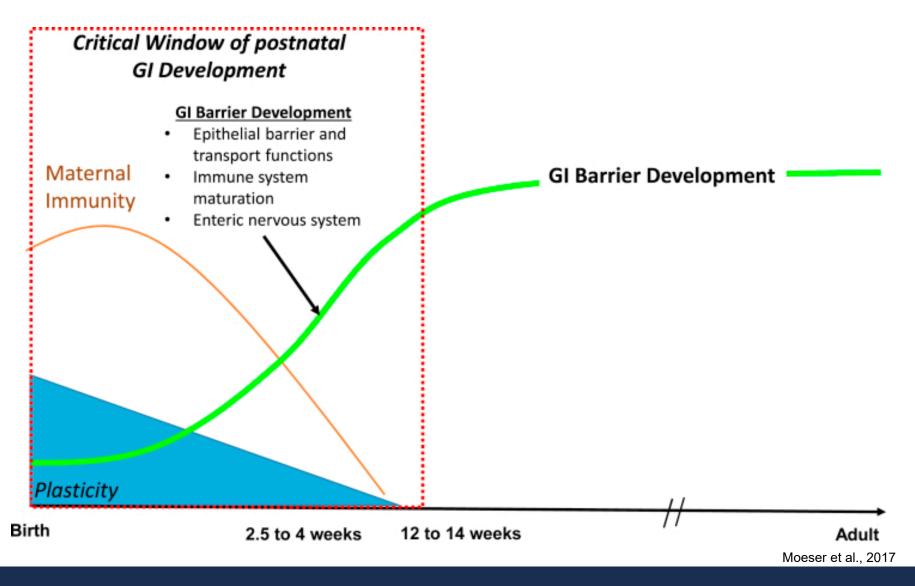
- Digestion and absorption of nutrients
- Physical barrier against pathogenic agents
- Large immune organ
- Nutrient chemo-sensing







Focus on the GUT of weaning pigs



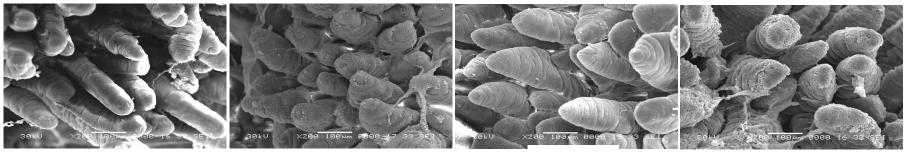
Weaning stress

- Maternal separation
- Environmental change
- Increased exposure to pathogens
- Social hierarchy stress
- Move to solid feed
- Transportation stress





Weaning stress on intestinal morphology

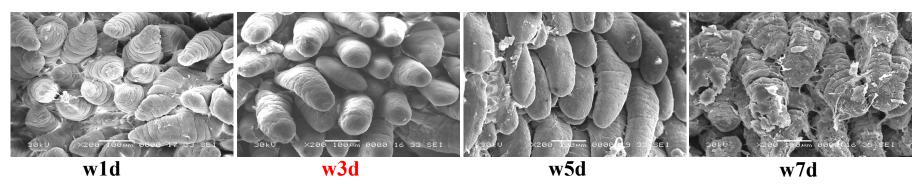


d1

d7

d14

d21

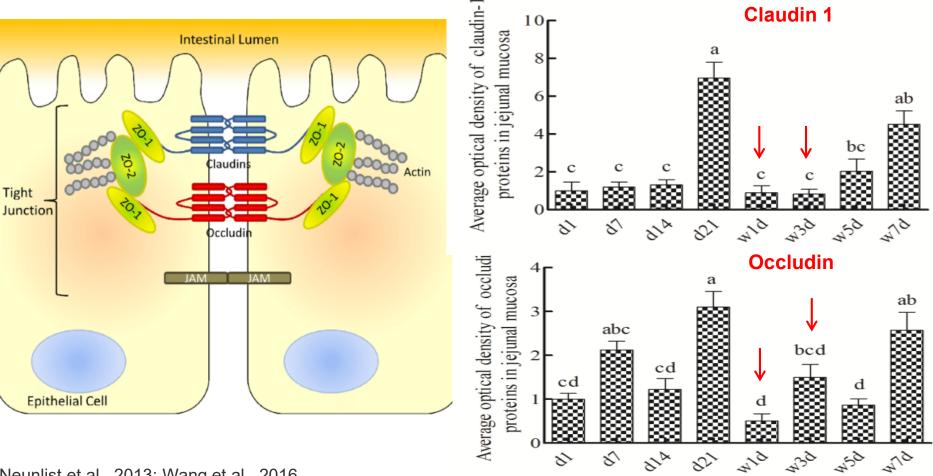


- Pre-weaning: d 1 to 21, villi surface was increased
- Post-weaning: reduced villi number and folding

Wang et al., 2016

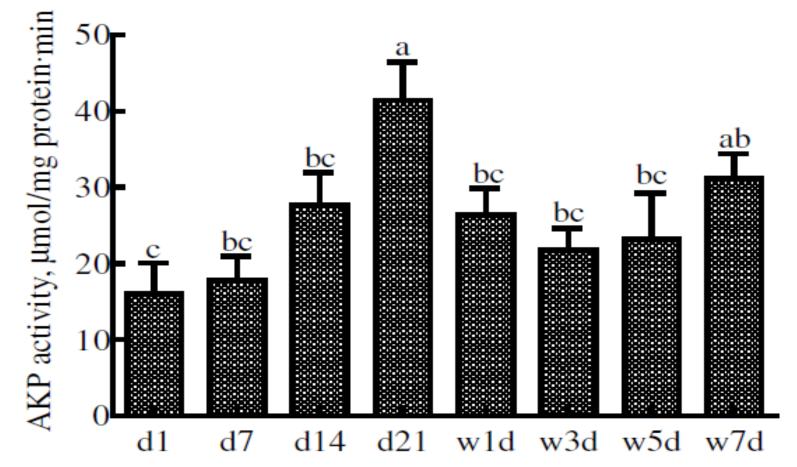


Weaning stress on intestinal barrier function



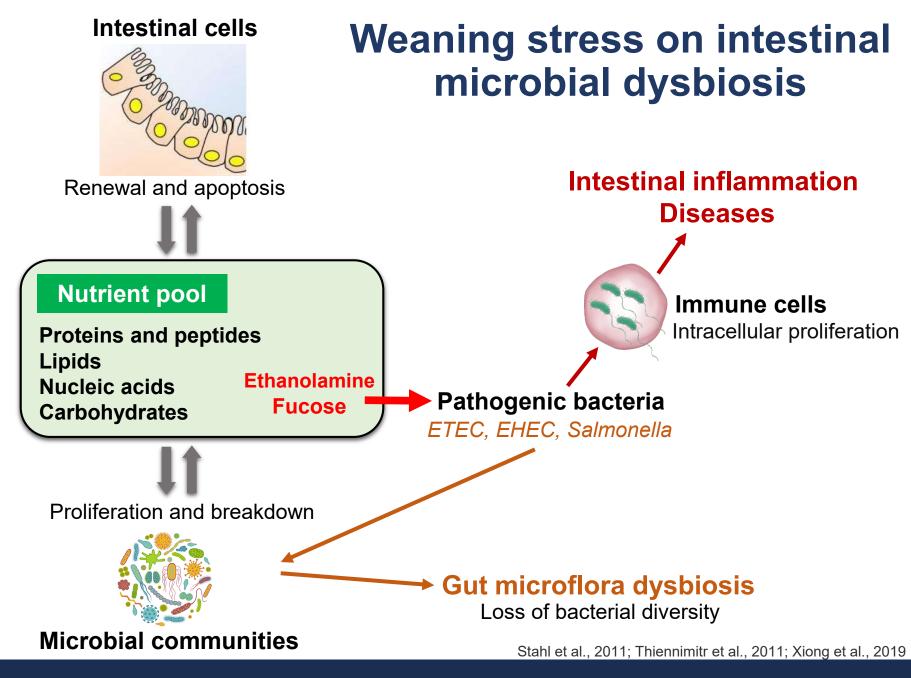
Neunlist et al., 2013; Wang et al., 2016

Weaning stress on intestinal barrier function, cont.

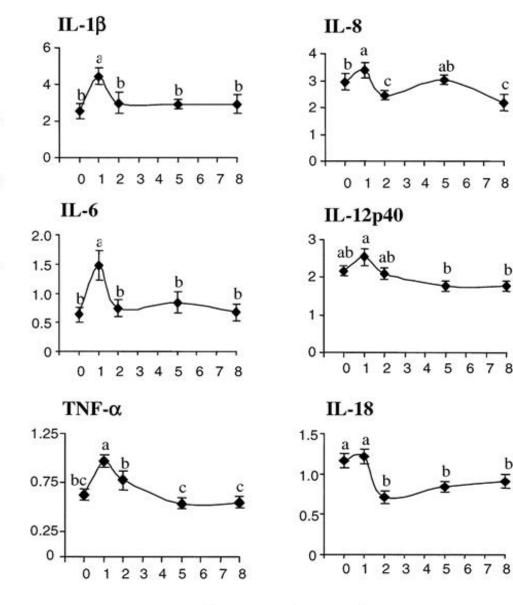


Wang et al., 2016





Middle of the small intestine



Normalized Values (arbitrary units)

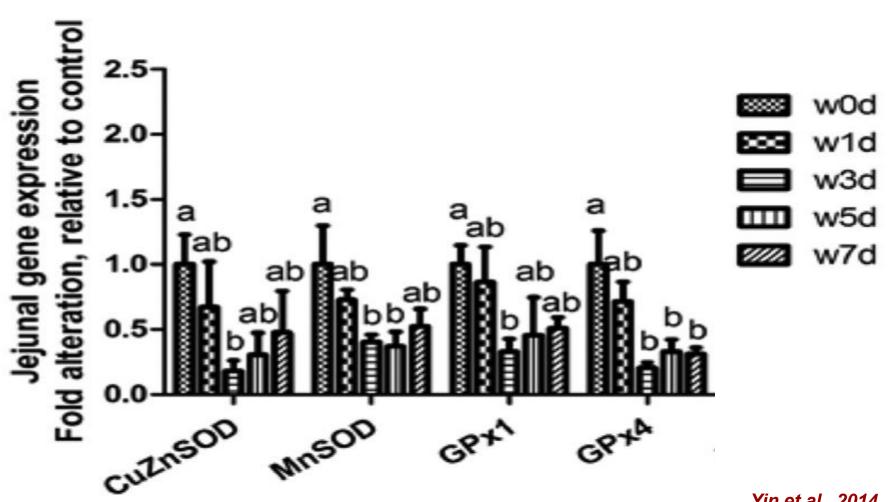
Weaning stress on intestinal mucosal immunity

- Weaning induces a transient gut inflammation in pigs
 - Enhanced proinflammatory cytokines
 - Increased intestinal CD4+ and CD8+ T lymphocytes
 - Up-regulated matrix metalloproteinase
 - Down-regulated MHC I expression
 - Reduced secretory IgA

Days post-weaning

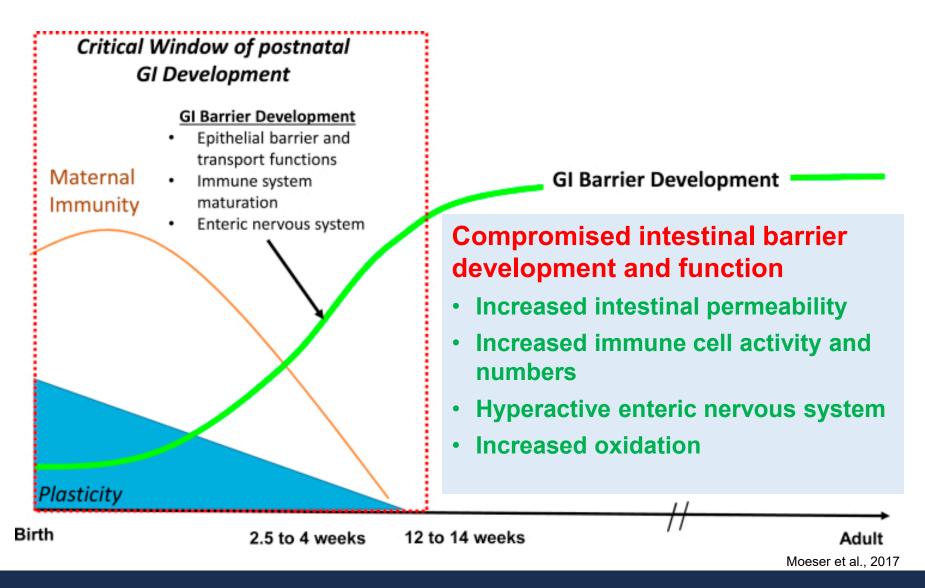
McCracken et al., 1999; Pié et al., 2004

Weaning stress on intestinal oxidative status



Yin et al., 2014

Focus on the GUT of weaning pigs



How to define a healthy gut

- Effective nutrient digestion and absorption
- Effective waste excretion
 Overall, should be concomitant with optimal performance

(the absence of diseases)

- A functional and protective gut immunity
- A minimal activation of stress/neural pathways

Pluske et al., 2018



Nutritional strategies

- Optimization of feed formulation
- Utilization of low protein diet in postweaning period
- Enhancement of feed processing and manufacturing
- Supplementation of feed additives



Feed additives

- Improvement of nutrient digestion and absorption (i.e. exogenous enzymes)
- Regulation gut microbiota to more favorable bacterial species (i.e. prebiotics & probiotics)
- Immune modulation to enhance disease resistance of weaned pigs (i.e. β-glucan, phytochemicals)





Two examples

Functional amino acids

Short chain fatty acids

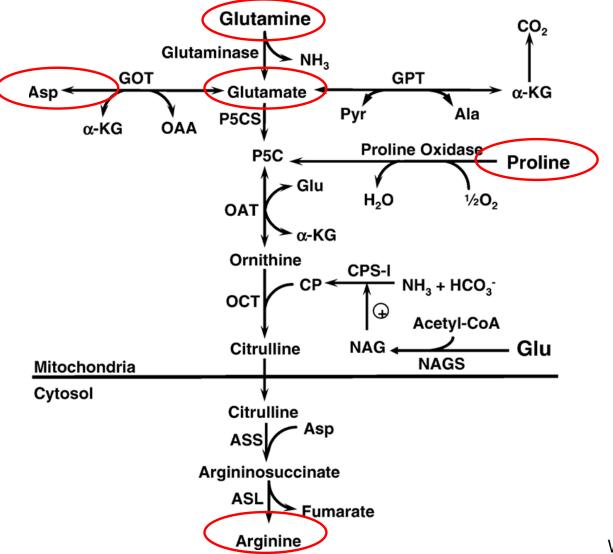


Functional amino acids

- Indispensable amino acids vs. dispensable amino acids
- Functional amino acids
 - Extra benefits to the host beyond the nutrient contribution
 - Arginine family (glutamate, glutamine, proline)
 - Aromatic amino acids (tryptophan, phenylalanine, tyrosine)



Arginine family



Wu et al., 2007

Arginine family

Substrates for tissue protein synthesis

Regulate

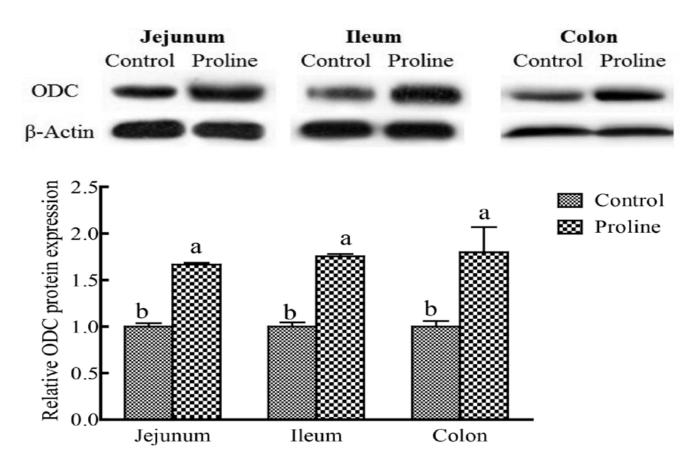
- Cellular signaling
- Hormone synthesis and secretion (insulin, glucagon, etc.)
- Endothelial function, vasodilation, blood flow
- Nutrient metabolism
- Intestinal integrity and function
- Immune function and health

Wu et al., 2007





Proline

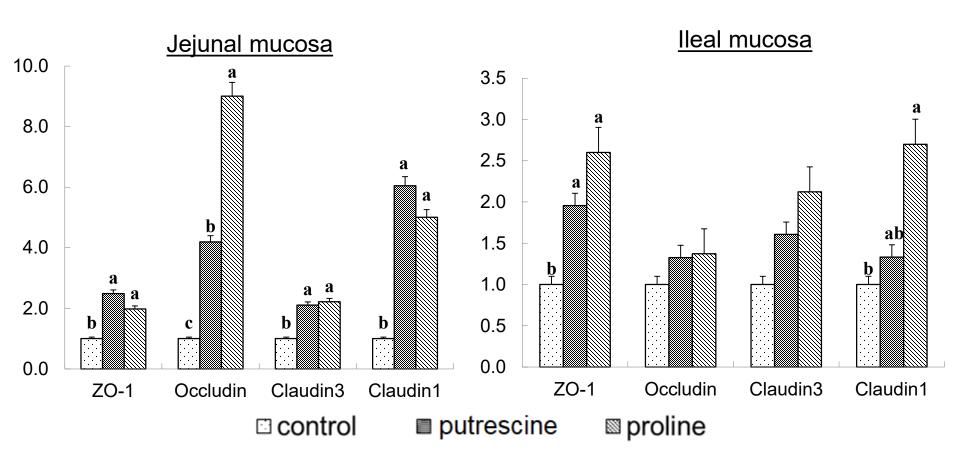


 Oral administration of proline enhanced protein expression of ornithine decarboxylase (ODC) activity in jejunum, ileum, and colon

Tan et al., 2017



Proline



 Oral administration of proline enhanced the expression of proteins involved in tight junction barrier of weaned pigs

Tan et al., 2017



Aromatic amino acids pig systemic immunity

Sorum na/ml	Saline		LPS	
Serum, pg/mL	Basal diet	TPT diet	Basal diet	TPT diet
IL1β	254°	215 ^c	1384 ^a	793 ^b
IL6	17.1 ^c	9.4 ^c	270 ^a	132 ^b
IL8	98 ^c	96 ^c	1076 ^a	674 ^b
IL12	115 ^c	102 ^c	497 ^a	310 ^b
GM-CSF	154 ^b	113 ^c	189 ^a	161 ^b
ΤΝFα	0.06 ^c	0.07 ^c	326 ^a	171 ^b
IL4	317 ^b	660ª	167°	291 ^b
TGFβ1	897 ^a	883 ^a	416 ^c	623 ^b

• Aromatic amino acids: Trp, Phe, Tyr, 1.5*NRC, 2012

Tan et al., 2017

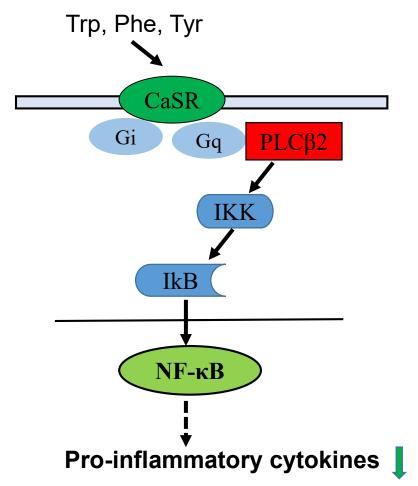
Aromatic amino acids pig intestinal immunity

Gene	Saline		LPS		
expression	Basal diet	TPT diet	Basal diet	TPT diet	
IL6	1 ^{ab}	0.76 ^b	1.41 ^a	0.44 ^b	
IL12	1 ^b	0.91 ^b	1.71 ^a	0.37 ^c	
IL18	1 ^{ab}	1.04 ^{ab}	1.52ª	0.47 ^b	
TNFα	1 a	1.10 ^a	1.28ª	0.27 ^b	
TGFβ	1 ^c	2.57 ^{ab}	1.93 ^b	3.10 ^a	

Tan et al., 2017



Aromatic amino acids Potential mechanisms



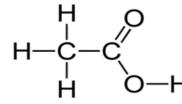
- Dietary supplemented with aromatic amino acids increased CaSR and PLCβ2 protein expression levels
- But decreased p-NF-κB, IKKα/β, and IκB protein expression levels in the LPS-challenged piglets

Tan et al., 2017

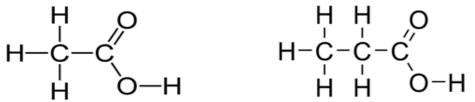


Short chain fatty acids

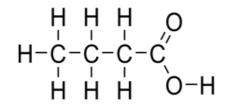
- Fatty acids with a chain of < 6 carbon atoms
 - Acetate, propionate, and butyrate
- Produced by microbial fermentation in the gastrointestinal tract of pigs
- Major fuel source for colonocytes (90% of butyrate)
- Derivatives: salts (Ca, Na), monobutyrin, tributyrin



Acetic acid (acetate)



Propionic acid (propionate)



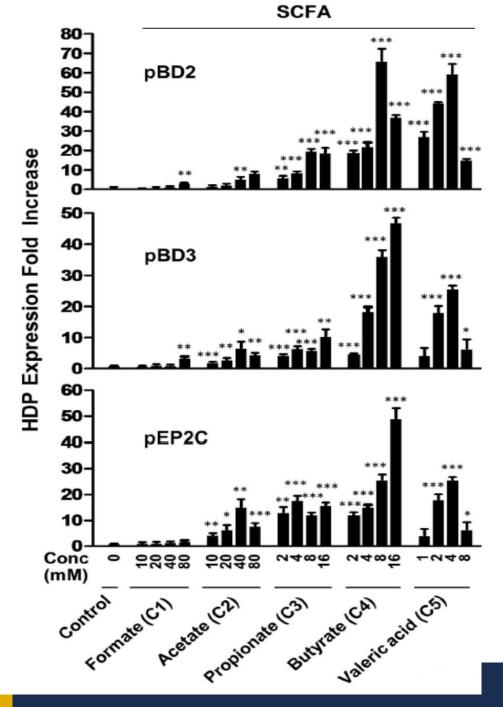
Butyric acid (butyrate)

Short chain fatty acids Antimicrobial effects of butyric acid

Gram-negative bacteria	MIC, mg/mL	Gram-positive bacteria	MIC, mg/mL
<i>E. coli</i> , wild type	2.3	Enterococcus faecalis	2.0
<i>E. coli</i> , F18	2.5	Clostridium perfringens	1.2
<i>Salmonella</i> Typhimurium, wild type	2.7	Streptococcus pneumonia	1.0
<i>Salmonella</i> Typhimurium, disease break	2.6	Streptococcus suis	0.7
<i>Campylobacter jejuni</i> , wild type	0.5	MIC: minimal inhibitory concentration	
<i>Campylobacter jejuni</i> , disease outbreak	0.7		

Kovanda et al., 2019





Short chain fatty acids

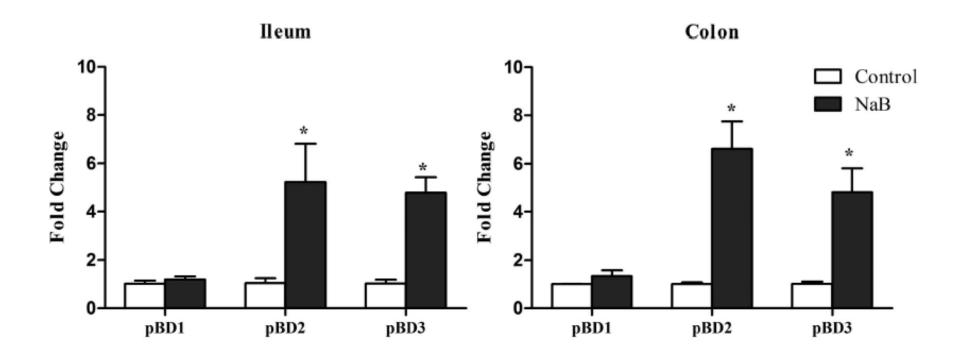
Host defense peptides, in vitro

- Also known as antimicrobial peptides
- Defensins or cathelicidins
- Small, positively charged, and amphipathic
- Disturb cell membrane structure, penetrate into cells, regulate intracellular pathways, cause bacterial cell death

Zeng et al., 2013

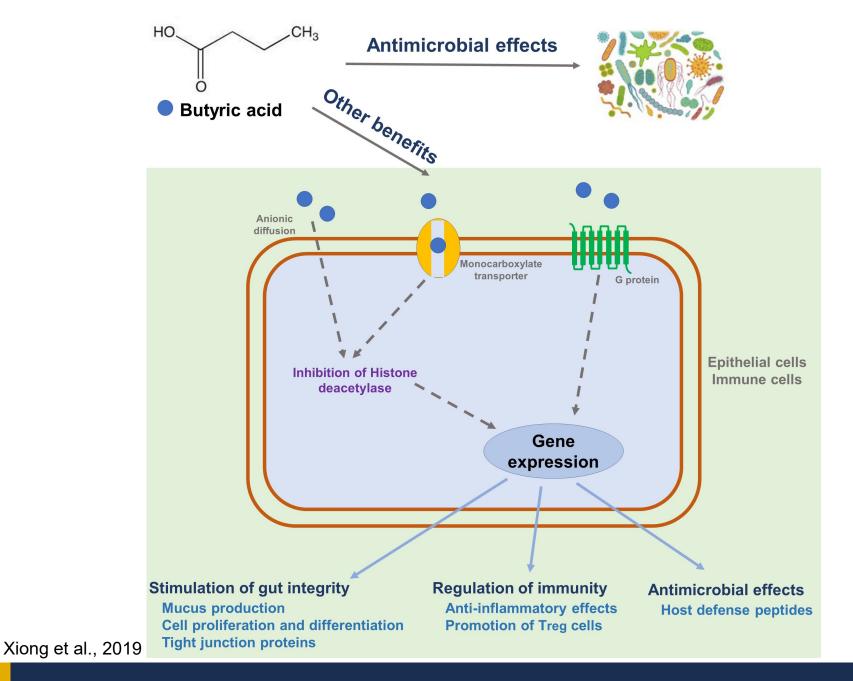


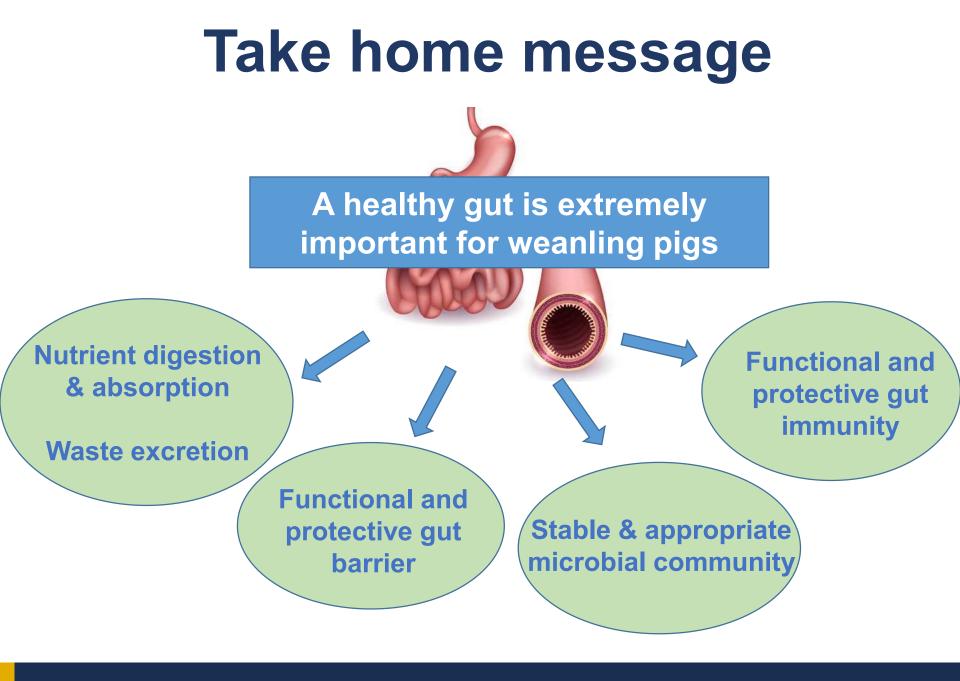
Short chain fatty acids Host defense peptides, in vivo



• Weaning pigs, 0.2% sodium butyrate, 10 days

Xiong et al., 2016





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